

09239902.0425010
Application
for
United States Letters Patent

To all whom it may concern:

Be it known that we

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have invented certain new and useful improvements in

A METHOD FOR REGULATING GENES WITH ELECTROMAGNETIC RESPONSE ELEMENTS

of which the following is a full, clear and exact description.

A METHOD FOR REGULATING GENES WITH ELECTROMAGNETIC
RESPONSE ELEMENTS

BACKGROUND OF THE INVENTION

Throughout this application, various publications are referenced to by arabic numerals within parentheses. Disclosures of these publications in their entireties are hereby incorporated by reference into this application to more fully describe the state of the art to which this invention pertains. Full bibliographic citations for these references may be found at the end of this application, preceding the claims.

Gene therapy was proposed approximately 20 years ago as a way to ameliorate genetic defects by providing a source for missing essential genetic components. The injection of copies of the gene responsible for the production of a specific protein directly into the targeted area by means of a viral vector was considered a mode of insuring that the protein required would be synthesized at the site where it was needed. This approach offered a distinct advantage over prior conventional treatment of metabolic diseases, which required continuous injection of gene product from exogenous sources.

The principle behind gene therapy is simple; however, practical application has been difficult. Failure of early gene therapy was mainly due to three problems. Firstly, there were difficulties in efficiently transducing primary quiescent human cells *in vivo*. Secondly, there were strong immune responses to the gene therapy vectors, as well as to the foreign therapeutic transgenes that rapidly eliminated trans-gene expressing cells in humans. Thirdly, there was an ability of many cell types to shut off the viral promoters that controlled transgene expression in humans.

TUESDAY, SEPTEMBER 28, 2004

5 One positive outcome of these early efforts of gene therapy was
the demonstration that introducing cloned genes into humans
could be safe, with little or no morbidity. More recently, new
vectors have been engineered, including adenoviruses and even
naked DNA, enhancing the efficiency of *in vivo* gene delivery
10 and reducing the immunogenicity of vectors and transgenes.

There is a need for a safer, more effective, and more precise
method of gene therapy.

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SUMMARY OF THE INVENTION

The present invention provides a unique method for gene regulation, using electromagnetic response elements. In the present invention, exogenously introduced genes, in gene therapy, are regulated by the introduction of electromagnetic response elements (EMREs) into the gene promoters that do not have them to serve as "switches." Exposure to electromagnetic fields of 8 μ T 60Hz for 30 minutes induces gene expression, because the switches make the gene now responsive to EM fields. The electromagnetic field response elements, therefore, are the "switches." The present invention therefore provides a non-invasive technique in gene therapy.

In this way, a safer, more effective, and more precise method for gene therapy is provided for inducing production of desired gene products. The present invention is therefore an improvement over the invasive character of current gene therapy protocols.

The electromagnetic field response elements, therefore, can be introduced into any gene promoter not having them. Examples are insulin and the cystic fibrosis gene. The electromagnetic field response elements can be introduced into any gene that would supply a missing gene product that the person does not already have due to some genetic consequence.

The present invention not only regulates and programs gene promoters to induce genetic information, but it does so in a patient-friendly manner.

Applying an electromagnetic field to the introduced gene containing the new electromagnetic field response elements induces gene expression.

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5 In summary, the present invention in one embodiment provides a
non-invasive method for gene regulation during gene therapy,
comprising the steps of: introducing electromagnetic field
response elements into a gene promoter not having any
electromagnetic field response elements to serve as switches
10 for regulating exogenously introduced genes; and applying an
electromagnetic field to the introduced electromagnetic field
response elements to induce gene expression.

15 The present invention in another embodiment provides a non-
invasive method for gene regulation during gene therapy,
comprising the steps of: introducing at least one
electromagnetic field response element into a gene promoter not
having any electromagnetic field response elements to serve as
switches for regulating exogenously introduced genes; and
applying an electromagnetic field to each introduced
electromagnetic field response element to induce gene
expression.

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BRIEF DESCRIPTION OF THE FIGURES

Fig. 1 illustrates construction of EMRE-expression vectors; and

Fig. 2 illustrates CAT and Luciferase activities, wherein samples in lane 1 were sham-exposed (30 mins), and samples in lanes 2, 3, and 4 were exposed to 8 μ T 60Hz EM fields (30 mins).

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DETAILED DESCRIPTION OF THE INVENTION

The present invention in one embodiment provides a non-invasive method for gene regulation during gene therapy, comprising the steps of: introducing electromagnetic field response elements into a gene promoter not having any electromagnetic field response elements to serve as switches for regulating exogenously introduced genes; and applying an electromagnetic field to the introduced electromagnetic field response elements to induce gene expression.

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The introduced electromagnetic field response elements may be nCTCTn sequences in an HSP70 gene promoter. A number of the nCTCTn sequences may be 3. The nCTCTn sequences may lie between about -230 and about -160 in the HSP70 gene promoter.

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The introduced electromagnetic field response elements may be nCTCTn sequences in a c-myc gene promoter. A number of the nCTCTn sequences may be 8. The nCTCTn sequences may lie between about -1257 and about -353 in the c-myc gene promoter.

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The electromagnetic field is preferably applied at a field strength of about 8 μ T and a frequency of about 60Hz for a time of about 30 minutes.

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The present invention in another embodiment provides a non-invasive method for gene regulation during gene therapy, comprising the steps of: introducing at least one electromagnetic field response element into a gene promoter not having any electromagnetic field response elements to serve as switches for regulating exogenously introduced genes; and applying an electromagnetic field to each introduced electromagnetic field response element to induce gene expression.

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5 Each introduced electromagnetic field response element may be
 an nCTCTn sequence in an HSP70 gene promoter. Each introduced
 electromagnetic field response element may be an nCTCTn
 sequence in a c-myc gene promoter. The electromagnetic field
 is preferably applied at a field strength of about 8 μ T and a
10 frequency of about 60Hz for a time of about 30 minutes.

15 Low frequency electromagnetic (EM) fields induce increased
 expression of the stress response gene HSP70 (6) (2). There
 are several parallels in the biochemical pathways induced by
 electromagnetic fields and heat shock, but there are striking
 differences as well. Both pathways involve the binding of heat
 shock factor 1 (HSF1) to a heat shock element (HSE), but
 regulation of HSP70 gene expression by electromagnetic fields
 involves three nCTCTn binding sites in the HSP70 promoter that
 lie between -230 and -160, upstream from the transcription
 initiation site. These three nCTCTn sequences appear to act as
 electromagnetic field response elements (EMREs), since the
 ability of an electromagnetic field to induce stress proteins
 gradually disappears as the EMREs are mutated one by one (7)
 (9). Removal of EMREs by mutation does not affect the response
 to heat shock, since the heat shock domain is downstream from
 the electromagnetic field domain in the HSP70 promoter, i.e.,
 between -106 and -67 (6) (8) (9).

30 A 900bp region in the c-myc promoter (-1257 to -353) is
 responsive to electromagnetic fields (5). Recent reanalysis of
 this 900bp region revealed eight nCTCTn sequences within this
 DNA fragment. These eight EMREs in the c-myc promoter could
 account for the electromagnetic field sensitivity of the c-myc
 gene, and the resultant increased c-myc transcript levels in
 cells exposed to electromagnetic fields (4).

5 To determine whether EMREs can serve as switches to regulate
exogenously introduced genes, the 900bp fragment of the *c-myc*
promoter was placed upstream of CAT (chloramphenicol
transferase) or luciferase reporter constructs that were
otherwise unresponsive to electromagnetic fields. EMRE-
10 reporter constructs were transfected into HeLa cells and
transflectants exposed to electromagnetic fields. Protein
extracted from EM field-exposed transflectants showed increased
CAT and luciferase activities, whereas no increase in CAT or
luciferase was measurable in the unexposed controls. Three
15 kinds of controls were used: transflectants that were sham-
exposed, transflectants lacking EMREs, and non-specific protein.
These data support the theory that EMREs can be inserted into
the promoters of exogenously introduced genes to serve as
switches that respond to electromagnetic fields. This would
provide a new and powerful non-invasive technique for
regulating gene expression during gene therapy.

MATERIALS AND METHODS

Cell culture and transfections

As previously described, HeLa cells were used for transient
transfactions and the lipofectin method (Gibco/BRL, Cat #
18292-011) was used for transfection as described (6) (7) (8).

900bp segment from the *c-myc* promoter

The 900bp region of the *c-myc* promoter containing eight copies
of nCTCTn extends from -353 (PVUII site) to -1257 (ClaI site).

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pAH-11-CAT HSP70 deletion construct

A diagrammatic representation of this construct is presented in
reference (9). This construct contains the first 111 base

5 pairs upstream from the transcription initiation site and includes the heat shock domain (-106 to -67). There are no nCTCTn binding sites in this construct and it is not responsive to electromagnetic fields (9).

10 Construction of EMRE-CAT expression vector

Fig. 1A illustrates construction of the EMRE-CAT expression vector (pΔ11+ 900bp + CAT). Plasmid pΔ11-CAT was digested with Hind III and PVU II, harvested from gel. Two oligonucleotides were used for PCR which allowed us to create two enzyme sites and amplify the 900bp region from *c-myc* promoter.

15 1. CCTGAGCTCTTCTTGATCAGAATCGATA
2. TCTAAGCTTCTTGATCAGAATCGATG

1 μ l of plasmid (digested with Hind III and PVUII) was mixed with 3 μ l PCR product, placed at 12°C overnight for ligation and transformed using DH52 bacteria. Clone hybridization verified insert.

20 Construction of EMRE- luciferase expression vector

Fig. 1B illustrates construction of the EMRE- luciferase expression vector (PGL₃ + 900bp + luciferase). A luciferase expression vector (PGL3 Promega) was digested with SacI and Smal and harvested from a gel. Two oligonucleotides (see above) were used for PCR, which allowed creation of two enzyme sites and amplified the 900bp region from *c-myc* promoter. 1 μ l of digested plasmid was mixed with 3 μ l PCR product, placed at 12°C overnight for ligation, and transformed using DH52 bacteria. Clone hybridization verified insert.

25 Protein

30 Protein was extracted and concentrations determined as previously described (6) (7) (8) (9).

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CAT assay

CAT assays were performed as previously described (6) (7). Results were quantified using a PhosphorImager and ImageQuant software.

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Luciferase assay

Luciferase activity was determined (Luciferase Assay Kit) (Promega #E1501) and results quantified as suggested by Promega.

Magnetic field exposures of transflectants

Transflectants were exposed and sham-exposed as previously described (7) (9).

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30Heat shock

Samples from cells that had been heat shocked (43°C) served as positive controls for CAT assay. Petri dishes containing transflectants were wrapped in Parafilm, placed in a mu metal box (to shield them from exposure to the magnetic fields generated by the water bath heating motor) and immersed in the water bath at 43°C for 30 minutes. Petri dishes were removed from the water bath and, following an additional 30 minutes at 37°C, protein was extracted (3) (6).

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Electromagnetic field exposure system

Two fully functional exposure units provided simultaneous sham and experimental exposures. Exposures used Helmholtz coils (Electric Research and Management, Pittsburgh, PA) that consisted of 19-gauge wire bundles wound 164 times around a square form 13cm long and 14cm wide with 8cm spacing. The coils were energized by a function generator (11 MHz Wavetek Stabilized Function Generator, model 21). A digital multimeter

5 was used to measure the field intensity and verify the systems
operation (Fluke 87 digital multimeter). Field parameters were
monitored with a Hitachi V-1065 100MHz oscilloscope and
calibrated inductive search coil (25X; Electro-Biology Inc.,
10 Parsippany, NJ). Detailed description of the exposure system,
including background magnetic fields in the incubator, harmonic
distortion, DC magnetic fields and mean static magnetic fields
in the incubator, both vertical and horizontal components, can
be found in reference (4). Cells were placed on a Plexiglas
15 stand in a horizontal orientation; i.e., the entire area of the
dish was exposed to the field. The bottom of the dish was 2cm
below the axis level. The height from dish bottom to top
surface of liquid was approximately 1.1 cm. The height of the
liquid was 0.6cm. The calculated electric field was ~11 μ V/m
15 for an 8uT exposure.

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Mu metal shielding

Helmholtz coils were enclosed within Mu metal containers to minimize stray fields during electromagnetic field exposures. Both active (experimental) and sham-exposed coils (controls) were enclosed in a 30 cm high, 15 cm diameter cylindrical mu metal container (.040" thickness) (Amuneal Corp. Philadelphia, PA). The 60Hz shielding factor is (Min.) 90.1 (39.08dB). Sham-exposed controls and experimental exposures are performed simultaneously in identical mu metal containers.

Statistical analyses

30 A sufficient number of experiments were performed to assure statistical significance. Statistical significance is determined by a multifactor analysis of variance program (INSTAT).

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Results

As will be discussed below, it was shown that EMREs increase luciferase activity in transfectants exposed to EM fields.

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To determine whether the nCTCTn sequences (EMREs) that are EM field responsive would confer EM field responsiveness to a reporter construct lacking these sequences, a 900bp region from the *c-myc* promoter containing eight copies of nCTCTn was ligated to a PGL3 plasmid containing a portion of the SV40 promoter and carrying the luciferase gene (see Fig. 1A). This plasmid construct was transfected into HeLa cells and the transfectants exposed to 8 μ T 60Hz fields for 30 minutes, followed by an additional 30 minutes out of the field prior to protein extraction for the luciferase assay. Luciferase activity increased an average of 61%.

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Fig. 2A is a bar graph showing luciferase activities. Fig. 2A illustrates: (1) luciferase activity in protein extracted from transfectants containing luciferase construct plus the 900bp insert (sham-exposed); (2) luciferase activity using non-specific protein (negative control) (EM field-exposed); (3) luciferase activity in protein extracted from transfectants containing luciferase construct minus the 900bp insert (EM field-exposed); and (4) luciferase activity in protein extracted from transfectants containing luciferase construct plus the 900bp insert (EM field-exposed).

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Therefore, three sets of controls were used: (1) sham-exposed transfectants that served as controls for electromagnetic field exposure, and showed no significant luciferase activity; (2) transfectants containing the luciferase reporter construct without the 900bp insert served as controls for background and

5 showed no measurable luciferase activity; and (3) non-specific protein served as negative controls with no measurable activity. These transfectants were not responsive to heat shock, as expected from the absence of heat shock consensus sequences (nGAA)n in this plasmid construct.

10 It was also shown that EMREs increase CAT activity in constructs exposed to EM fields. In similar experiments with a CAT reporter construct, the 900bp region from the c-myc promoter containing eight nCTCTn was ligated to pΔ11- CAT (see Fig. 1B), transfected into HeLa cells, and the transfectants exposed to an 8μT 60Hz field for 30 minutes, followed by an additional 30 minutes out of the field prior to protein extraction for the CAT assay. There was an average 60% increase in CAT activity. The same three sets of controls described above were employed in these experiments (see Fig. 15 2B): (1) sham-exposed transfectants served as controls for electromagnetic field exposure, and showed no significant CAT activity; (2) transfectants containing the CAT reporter construct without the 900bp insert (pΔ11-CAT) served as controls for background; protein extracts from these transfectants showed no measurable CAT activity; and (3) non-specific protein served as negative controls. Transfectants with and without the 900bp insert were heat shocked for 30 minutes at 43°C followed by protein extraction after an additional 30 minutes out of the heat. There was an average 20 25 30 45% increase in CAT activity in heat shocked transfectants. The pΔ11 plasmid contains the heat shock domain, -106 to -67 and therefore response to heat shock served as an additional control.

35 Therefore, Fig. 2B illustrates: (1) CAT activity in protein from transfectants containing CAT construct plus the 900bp

5 insert (sham-exposed); (2) CAT activity using non-specific
protein (negative control) (EM field-exposed); (3) CAT activity
in protein from transfectants containing CAT construct minus
the 900bp insert (EM field-exposed); and (4) CAT activity in
protein from transfectants containing CAT construct plus the
10 900bp insert (EM field-exposed).

Discussion

15 Because electromagnetic fields penetrate tissues without
attenuation, they must penetrate to the cell nucleus with its
DNA and interact with moving charges there (1). There are
conducting electrons in DNA (12), and direct measurements of
electrical transport through DNA have been made. The dynamics
of DNA-mediated electron transfer at the femtosecond level have
been measured (14). Conduction in DNA appears to depend on
specific structure, since different DNA sequences have
different conductivities (10). Therefore, electromagnetic
fields could theoretically interact preferentially with
specific DNA sequences, and the nCTCTn sequences (EMREs) in the
HSP70 and *c-myc* promoters used in these studies may be such
sequences.

25 We have shown that these sequences are critical for
electromagnetic field responsiveness in our experiments, and
other data appear to support this. In totally unrelated
investigations, one study showed that low frequency
electromagnetic field stimulation in nigro-striatal lesioned
30 rats with chromaffin transplants induced changes in the
subventricular zones and led to significant motor improvements
in a rat Parkinson model (13). A second report from the same
laboratory has used differential display to analyze possible
alterations in DNA of electromagnetic field-exposed chromaffin
35 cells.

5 Differential bands observed in the EM field-exposed group show
changes in gene expression induced by electromagnetic fields.
One specific differential band in the EM field-exposed samples,
containing 349bp, was sequenced. In an independent analysis of
10 this DNA fragment, we have identified three copies of the
electromagnetic field response element (nCTCTn) that we
described herein. A computer search may determine whether this
349bp DNA fragment is contained in the promoters of any known
genes, possibly a specific gene related to the differentiation
process of chromaffin cells.

15 Electromagnetic fields induce gene expression (2) (9) and
activation of the gene by electromagnetic fields requires
specific EMREs, which control genes when placed upstream of
reporter constructs. Their ability to confer electromagnetic
field responsiveness suggests the use of EMREs in the control
and regulation of gene therapy. The characterization of a
cellular promoter system that can be regulated, such as
described here, provides a novel, noninvasive, technique for
the regulation of transgene expression in humans without
interfering with normal physiologic function. The applied
electromagnetic field can be directed to the region where the
gene product is needed, and, since the electromagnetic field
intensities needed to affect EMREs are well below the human
perception threshold, their introduction and presence would not
30 be felt by the patient.

35 An example of such application would be the introduction of an
exogenous insulin gene containing one or more EMREs placed
upstream of the gene. Regulation would be provided by the
simple and safe application of electromagnetic fields. The
whole operation would be made automatic by having the EM field
generating circuit activated by an implanted glucose sensor

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5 responsive to pre-set blood glucose levels.

Our results show that the eight nCTCTn sequences (EMREs) in the 900bp DNA fragment from the *c-myc* promoter are effective in regulating CAT or luciferase activity. However, not all eight EMREs may be needed for a response (9). The EM-induced expression of HSP70 is mediated through three EMREs in the 10 human HSP70 promoter. Electromagnetic field exposure of HSP70 promoter constructs, linked to a CAT reporter gene and containing all three sites, showed more than a three fold increase in CAT activity. Yet, the presence of even one site was sufficient for a 1.5 fold increased CAT response. These 15 data show that even a single EMRE can promote interaction with electromagnetic fields. The data also suggest that the level of interaction appears to be roughly proportional to the number of EMREs.

According to an embodiment of this invention, then, nCTCTn 20 sequences, taken from the *myc* promoter, were attached to HSP70 constructs that didn't contain them. The HSP70 promoter has three nCTCTn sequences in the electromagnetic field domain (230-160), but none in the heat shock domain (111-67). When 25 the nCTCTn sequences are inserted into the heat shock domain (which was previously responsive only to heat and not to electromagnetic fields) this promoter construct that previously did not respond to EM fields, now does respond and induces gene expression.

30 In this way, electromagnetic field response elements, i.e., nCTCTn sequences from the *c-myc* promoter, are actively incorporated into the HSP70 promoter and regulate and program gene expression; thus, inserting these nCTCTn sequences into a reporter construct (CAT or Luciferase) that was previously

5 unresponsive to EM fields, renders the gene electromagnetic field-responsive, and induces the gene activity.

Summary

10 A 900 base pair segment of the *c-myc* promoter, containing eight nCTCTn sequences, induces *c-myc* expression by electromagnetic fields. Similarly, a 70bp region of the HSP70 promoter, containing three nCTCTn sequences, induces HSP70 expression by electromagnetic fields. Removal of the 900 base pair segment of the *c-myc* promoter eliminates the ability of electromagnetic fields to induce *c-myc* expression. Similarly, removal of the 15 70bp region of the HSP70 promoter, with its three nCTCTn sequences, eliminates the response to electromagnetic fields. The nCTCTn sequences apparently act as electromagnetic field response elements (EMREs). To test whether introducing EMREs imparts the ability to respond to applied electromagnetic fields, the 900bp segment of the *c-myc* promoter (containing eight EMREs) was placed upstream of CAT or luciferase reporter constructs that were otherwise unresponsive to electromagnetic fields. EMREs-reporter constructs were transfected into HeLa cells and exposed to 8 μ T 60Hz fields. Protein extracts from EM field-exposed transfecants had significant increases in activity of both CAT and luciferase, compared with identical transfectants that were sham-exposed. Transfectants with CAT or luciferase constructs lacking EMREs remained unresponsive to 20 EM fields; that is, there was no increase in either CAT or luciferase activity. These data support the idea that EMREs can be used as switches to regulate exogenously introduced 25 genes in gene therapy.

30 Although embodiments of the invention have been described herein, numerous variations and modifications will occur to 35

5 those skilled in the art without departing from the scope of
the invention. The invention is not limited to the embodiments
disclosed, and is defined only by way of the following claims.

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